

SYNTHESES OF CYCLOTETRAPEPTIDES, AM-TOXIN ANALOGS, CONTAINING α -HYDROXYALANINE

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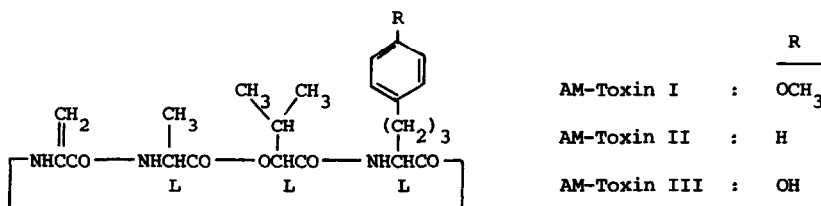
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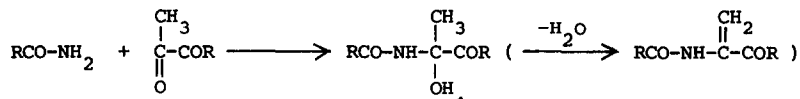
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Summary: Condensation of amide with α -keto acid to yield a α -hydroxyalanine (α -Hyla) or dehydroalanine residue was applied to syntheses of analogs of AM-toxins, cyclotetradepsipeptides *Cyclo*($-\alpha$ -Hyla-L-Ala-L-Val-L-Phe-), *cyclo*($-\alpha$ -Hyla-L-Ala-L-Hmb-L-Phe-) and *cyclo*($-\alpha$ -Hyla-L-Ala-L-Hmb-L-Tyr-) (Hmb, 2-hydroxy-3-methylbutanoic acid) were obtained from the corresponding pyruvyl-tripeptide amides in good yields by the treatment of anhydrous hydrogen fluoride.

AM-Toxins are host-specific phytotoxic metabolites produced by *Alternaria mali*, and cause veinal necrosis on apple leaves.¹⁾ The primary structures of three congeners, AM-toxins I, II and III, were determined to be cyclotetradepsipeptide.^{2,3)} In previous papers,⁴⁻⁷⁾ we reported the syntheses of AM-toxins I, II and their several analogs, and partly clarified the structure-activity relationship of the toxins. The synthetic methods used for AM-toxins I and II, in which dehydroalanine (Δ Ala) residue was formed by β -elimination of Ser residue, can

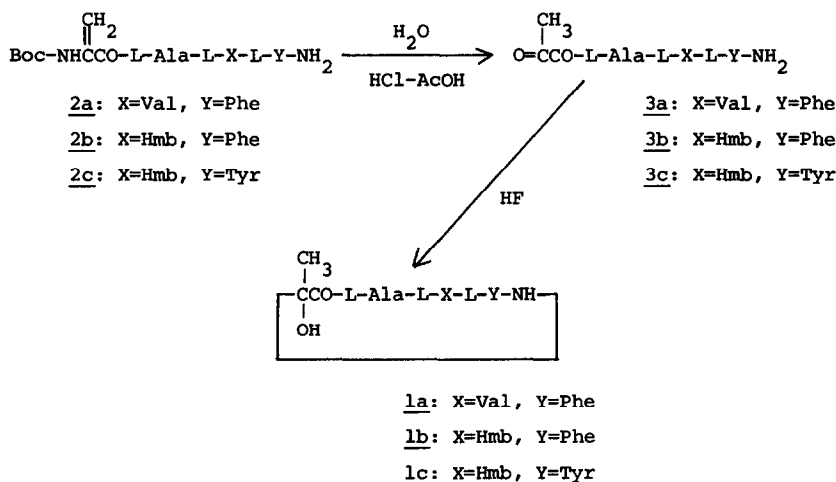


hardly be utilized for AM-toxin III because it contains a free hydroxy group. Therefore, we tried to apply the condensation of amide with α -keto acid (Scheme 1). The synthesis of peptides



Scheme 1. Condensation of acylamide with α -keto acid.

through the condensation has been reported in the case of the synthesis of 2,5-piperadinediones.⁸⁾ Gross and Morell observed that the reversible reaction of degradation-formation of α,β -unsaturated amino acids takes place in the peptide antibiotics, nisin and subtilin.⁹⁾ Recently, Noda and Gross obtained a cyclic hexapeptide containing α -Hyalal or Δ Ala residue through the intramolecular condensation of pyruvyl-pentapeptide amide with C-terminal sequence of luteinizing hormone-releasing hormone by the action of anhydrous HF.¹⁰⁾ This paper describes the syntheses of AM-toxin analogs (1a, 1b and 1c) by the intramolecular condensation of pyruvyl-tripeptide amides (Scheme 2). As the first step of the experiments, three Boc- Δ Ala-tripeptide amides, namely Boc- Δ Ala-L-Ala-L-Val-L-Phe-NH₂ (2a), Boc- Δ Ala-L-Ala-L-Hmb-L-Phe-NH₂ (2b) and Boc- Δ Ala-L-Ala-L-Hmb-L-Tyr-NH₂ (2c), were prepared by the conventional method, and converted to the corresponding pyruvyl-tripeptide amides, namely Pyr-L-Ala-L-Val-L-Phe-NH₂ (3a), Pyr-L-Ala-L-Hmb-L-Phe-NH₂ (3b) and Pyr-L-Ala-L-Hmb-L-Tyr-NH₂ (3c) (Pyr, pyruvyl). Then, each of 3a-3c was treated with anhydrous HF to yield a cyclic peptide.



Scheme 2. Syntheses of AM-toxin analogs by the intramolecular condensation of pyruvyl-tripeptide amides.

Synthesis of 2a was performed in a stepwise manner from its C-terminus. Boc-L-Val-L-Phe-OMe (4) (mp 104-106°, $[\alpha]_D^{25}$ -26° in EtOH) was prepared by the coupling of Boc-L-Val-OH and H-L-Phe-OMe·HCl using 1-ethyl-3-(3'-dimethylaminopropyl)carbodiimide (WSCl) in yield of 74%. Removal of the Boc-group of 4 with 2 M HCl in ethyl acetate and coupling with Boc-L-Ala-OH using WSCI led to Boc-L-Ala-L-Val-L-Phe-OMe (5) (86%, mp 140-141°, $[\alpha]_D^{25}$ -42.5° in EtOH). The protected tripeptide 5 was converted to Boc-L-Ala-L-Val-L-Phe-NH₂ (6) (mp 210°, $[\alpha]_D^{25}$ -26° in EtOH) by the action of saturated ammonia in MeOH in yield of 95%. Action on 6 with HCl in ethyl acetate and coupling with Boc-ΔAla-OH, which was prepared from Boc-L-Ser(Tos)-OMe by the action of NaOH,¹¹⁾ using WSCI afforded 2a (49%, mp 185-187°, $[\alpha]_D^{25}$ -7.5° in EtOH).

Syntheses of 2b and 2c were initiated with Boc-L-Ala-L-Hmb-OH,⁵⁾ which was coupled with H-L-Phe-NH₂·HCl or H-L-Tyr-NH₂·HCl by WSCI to give Boc-L-Ala-L-Hmb-L-Phe-NH₂ (7) (55%, mp 120-122°, $[\alpha]_D^{25}$ -57° in EtOH) and Boc-L-Ala-L-Hmb-L-Tyr-NH₂ (8) (43%, mp 144-145°, $[\alpha]_D^{25}$ -57° in EtOH), respectively. Removal of the Boc-group of 7 or 8 and coupling with Boc-ΔAla-OH by WSCI led to 2b (68%, mp 99-100°, $[\alpha]_D^{25}$ -36° in EtOH) and 2c (40%, mp 130-133°, $[\alpha]_D^{25}$ +32° in EtOH).

The ΔAla-peptide amide 2a was converted to 3a by the treatment with 1 M HCl in AcOH in the presence of an equivalent amount of water,¹²⁾ and, without further purification, 3a was allowed to react with anhydrous HF for 1 h at 0°. After evaporation *in vacuo*, the residual solid was collected with the aid of ether. The compound obtained showed a major spot (R_f 0.57) and a minor spot (R_f 0.05) on silica gel TLC with the solvent system of chloroform-MeOH-AcOH (85:10:5). After purification by column chromatography using silicic acid and the solvent system of chloroform-dioxane (1:1), a single product was obtained in high yield (85%) (R_f 0.57, mp 199-202°, $[\alpha]_D^{25}$ -14.5° in DMF). By chemical and physical analyses, the compound was identified to be *cyclo*(-α-Hyala-L-Ala-L-Val-L-Phe-) (1a): *anal* (C₂₀H₂₈O₅N₄) C,H,N; mol wt¹³⁾ 404 (calcd 404); ¹NMR¹⁴⁾ (DMSO) δ 2.32 (s, CH₃), δ 7.04 (s, C^α-OH), δ 7.30 (s, NH) (signals arising only from α-Hyala residue are given). No dimer was detected. The treatment of 2a with anhydrous HF afforded the same product 1a (78%). Although the α-Hyala residue in 1a involves an asymmetric carbon atom, the α-Hyala residue was determined to have a single optical configuration of either L or D from the NMR data on 1.

The cyclic peptides 1b and 1c were also obtained from 3b and 3c, respectively, by the same procedure described above. Peptide 1b: yield, 74%; mp 167-169°; $[\alpha]_D^{25}$ -48° in EtOH; *anal* (C₂₀H₂₇O₆N₃) C,H,N; mol wt 405 (calcd 405); ¹NMR (DMSO) δ 2.36 (s, CH₃), δ 7.07 (s, C^α-OH), δ 7.30 (s, NH). Peptide 1c: yield, 77%; mp 151-153°; $[\alpha]_D^{25}$ -26.5° in EtOH; *anal* (C₂₀H₂₇O₇N₃) C,H,N;

mol wt 421 (calcd 421); ^1NMR (DMSO) δ 2.35 (s, CH_3), δ 7.07 (s, $\text{C}^\alpha\text{-OH}$), δ 7.26 (s, NH).

None of the synthetic analogs, la, lb and lc, showed the toxic activity for induction of necrosis on apple leaves. The present results, however, provide a facile method for cyclization of peptides, which gives only monomer in high yield containing an interesting amino acid residue, α -hydroxyalanine, with a definite optical configuration.

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References and Notes

- 1) T. Ueno, Y. Hayashi, T. Nakashima, H. Fukami, H. Nishimura, K. Kohmoto and A. Sekiguchi, *Phytopathology*, 65, 82 (1975).
- 2) T. Ueno, T. Nakashima, Y. Hayashi and H. Fukami, *Agric. Biol. Chem.*, 39, 1115, 2081 (1975).
- 3) T. Okuno, Y. Ishita, K. Sawai and T. Matsumoto, *Chemistry Lett.*, 635 (1974).
- 4) Y. Shimohigashi, S. Lee, T. Kato, N. Izumiya, T. Ueno and H. Fukami, *Agric. Biol. Chem.*, 41, 1533 (1977).
- 5) Y. Shimohigashi, S. Lee, H. Aoyagi, T. Kato and N. Izumiya, *Int. J. Pept. Protein Res.*, 10, 197 (1977).
- 6) Y. Shimohigashi, S. Lee, H. Aoyagi, T. Kato and N. Izumiya, *Int. J. Pept. Protein Res.*, 10, 323 (1977).
- 7) Y. Shimohigashi and N. Izumiya, *Int. J. Pept. Protein Res.*, 12, 7 (1978).
- 8) C. Shin, K. Sato, A. Ohtsuka, K. Mikami and J. Yoshimura, *Bull. Chem. Soc. Jpn.*, 46, 3876 (1973); J. Hausler and U. Schmidt, *Chem. Ber.*, 107, 2804 (1974); B. W. Bycroft and G. R. Lee, *J. Chem. Soc. Chem. Commun.*, 988 (1975).
- 9) E. Gross and J. L. Morell, *FEBS Lett.*, 2, 61 (1968).
- 10) K. Noda and E. Gross, unpublished data.
- 11) I. Photaki, *J. Am. Chem. Soc.*, 85, 1123 (1963).
- 12) J. Greenstein and M. Winitz, "Chemistry of the Amino Acids", Vol. 2, p. 843, John Wiley and Sons, Inc., New York, N.Y. (1961); E. Gross and J. L. Morell, *J. Am. Chem. Soc.*, 93, 4634 (1971).
- 13) Mol wt was determined on Hitachi RMS-4 mass spectrometer.
- 14) Recorded on JEOL JNM PS-100 spectrometer.

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